

Alzheimer Diseases

Soheila Madadi¹; Mehdi Mehdizaded^{2,*}

¹Department of Anatomy, Faculty of Medicine, Iran University of Medical Sciences, Tehran, IR Iran

²Cellular and Molecular Research Center, Department of Anatomical Sciences, Faculty of Advanced Technology in Medicine, Iran University of Medical Sciences, Tehran, IR Iran

*Corresponding author: Mehdi Mehdizaded, Cellular and Molecular Research Center, Department of Anatomical Sciences, Faculty of Advanced Technology in Medicine, Iran University of Medical Sciences, Tehran, IR Iran. Tel/Fax: +98-21-886622689, E-mail: mehdizadeh.m@iums.ac.ir

Received: April 26, 2014; Revised: May 18, 2014; Accepted: May 20, 2014

Keywords: Alzheimer Diseases; Amyloid beta-Peptides; Memory

Alzheimer's disease (AD), the most common cause of dementia is accompanied by progressive memory loss and other cognitive functions. The condition is estimated to affect approximately 36 million people, worldwide (1). AD is characterized by the presence of extracellular amyloid β ($A\beta$) deposits, intracellular neurofibrillary tangles and senile plaques in the cortex, hippocampus, basal forebrain and amygdala (2). Neurofibrillary tangles formation is the result of intracellular fibrillar aggregation of the microtubule-associated protein tau that is hyperphosphorylated and oxidized. Senile plaques consist of insoluble fibrillar $A\beta$. It is established that $A\beta$ is formed after sequential cleavage of amyloid precursor protein and secreted to the extracellular space. It also inhibits hippocampal long-term potentiation and disrupts the synaptic plasticity (3). In addition, $A\beta$ accumulation induces an elevation in levels of reactive oxygen species (ROS) in neurons, leading to apoptotic neuronal death in rats and mice (4). Studies showed that the accumulation of $A\beta$ in brain plays an important role in the pathophysiology of AD and a close correlation exists between $A\beta$ procedure and the neurodegeneration process of AD (5). There exists evidence suggesting that memory impairment in AD begins with changes in hippocampal synaptic functions and then gradually progresses to neurodegeneration and neuronal loss in these patients (6).

The $A\beta$ -induced damage in hippocampus might underlie some of the AD behavioral deficits. Long-term potentiation (LTP) is one of the most important forms of synaptic plasticity, linked to learning and memory (7). The $A\beta$ makes changes in LTP, in the hippocampus and consequently impairs cognition and memory in rodents (3) and is widely reported to cause lipid peroxidation in brain cell membranes, leading to 4-hydroxy-2-nonenal (HNE) and acrolein formation, both toxic to neurons. These products alter the membrane protein conformation and eventually lead to neuronal death (8). The $A\beta$ initiates free radical processes, resulting in protein ox-

idation, lipid peroxidation, ROS formation and cellular dysfunction, leading to calcium ion accumulation and subsequent neuronal death (9).

Pervious experimental studies have shown that $A\beta$ (25-35) induce a wide pattern of central modifications, reminiscent of the human physiopathology, particularly short- and long-term memory deficit, oxidative stress, apoptosis, neuroinflammation, acetylcholine impairment, hippocampus alteration, tau hyperphosphorylation and amyloid burden (10). The deposition of β -amyloid protein in brain is related to learning impairment and cholinergic neuronal degeneration and the β -amyloid protein-treated rats could be used as AD animal models (11). The key brain regions, involved in the Morris water maze (MWM) task navigation, include the striatum, the frontal lobe and especially, the hippocampus (12).

The hippocampus structure has a key role in cognition and psychological function. Animal studies have shown that this structure is rapidly and extremely affected by an $A\beta$ fragment injection ($A\beta$ (25-35)) in rats, damaging the structure and function of the hippocampus (13, 14). The hippocampus plays an important role in contextual memory; the hippocampus injuries negatively affect the MWM task performance (15).

Nitta et al. showed that the water maze task performance was impaired in β -amyloid-treated rats, and the choline acetyl transferase activity significantly decreased in the frontal cortex and hippocampus (16). Therefore, the β -amyloid protein deposition in brain is believed to be related to learning impairment and cholinergic neuronal degeneration. It also means that β -amyloid protein-treated rats could be used as animal models for AD (11). Moreover, the studies indicated that intracerebroventricular (ICV) injection of $A\beta$ (25-35), induced impairment in the passive-avoidance and radial-arm maze tasks, in the rat (11). Maurice confirmed the negative effect of ICV injection of $A\beta$ (25-35) on learning in the Y-maze, passive avoidance and water maze tasks (17). The studies also reported

that bilateral injection of A β (25-35) induced learning deficits in passive-avoidance tasks, in rats (18). The hippocampus has an important role in spatial navigation and consolidation of information from short-term to long-term memory. Evidence have suggested that the memory impairment in AD begins with changes in hippocampal synaptic functions and progresses to neurodegeneration and neuronal loss in these patients (19). It has been reported that A β administration makes changes in LTP in the hippocampus and consequently leads to cognitive dysfunction and memory impairment in rodents (20). It is clear that oxidative stress plays a role in AD-induced neurotoxicity in the brain. The A β enters the bilayer neuronal membrane and generates oxygen-dependent free radicals, causing lipid and protein oxidation (21). Oxidative stress disrupts the blood brain barrier, leading to toxic substances passage to the brain and ultimately, resulting in the progression of various neurodegenerative diseases. Furthermore, the A β deposition activates the acute immune response of microglial cells and astrocytes, leading to production and activation of inflammation-related proteins, including complement factors and cytokines like interleukin-1, interleukin-6 and tumor necrosis factor- α and therefore leading to synaptic damage, neuronal loss and the activation of other inflammatory participants (9, 22, 23). As mentioned above, oxidative stress, following A β , involves development and progression of the AD. Brain is sensitive to oxidative stress, due to low antioxidant and cell membrane lipid levels (24). Oxidative stress reflects an imbalance between the systemic ROS manifestation and a biological system ability of detoxifying the reactive intermediate molecules or easily repairing the resulting damage.

Therefore, the use of an external antioxidant is one of the most common therapeutic strategies for neurotoxicity treatment. Several experimental studies have shown that dietary enrichment with nutritional antioxidant could improve brain damage and cognitive function (25-28).

A great number of different spices and aromatic herbs have been used as antioxidants in neurological diseases (29, 30).

Antioxidants that prevent the detrimental consequences of A β are consequently considered to be a promoting approach to brain neuroprotection in the AD (31).

Acknowledgements

We would like to thank Farzaneh Hosseini for typing the manuscript.

Authors' Contributions

S. Madadi gathered and wrote the manuscript and M. Mehdizadeh edited it.

References

- Walsh DM, Teplow DB. Alzheimer's disease and the amyloid beta-protein. *Prog Mol Biol Transl Sci.* 2012;107:101-24.

- Mattson MP, Maudsley S, Martin B. A neural signaling triumvirate that influences ageing and age-related disease: insulin/IGF-1, BDNF and serotonin. *Ageing Res Rev.* 2004;3(4):445-64.
- Shankar GM, Li S, Mehta TH, Garcia-Munoz A, Shepardson NE, Smith I, et al. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med.* 2008;14(8):837-42.
- Allanbutterfield D, Castegna A, Lauderback C, Drake J. Evidence that amyloid beta-peptide-induced lipid peroxidation and its sequelae in Alzheimer's disease brain contribute to neuronal death. *Neurobiology of Aging.* 2002;23(5):655-64.
- Boncristiano S, Calhoun ME, Kelly PH, Pfeifer M, Bondolfi L, Stalder M, et al. Cholinergic changes in the APP23 transgenic mouse model of cerebral amyloidosis. *J Neurosci.* 2002;22(8):3234-43.
- Klyubin I, Ondrejcek T, Hayes J, Cullen WK, Mably AJ, Walsh DM, et al. Neurotransmitter receptor and time dependence of the synaptic plasticity disrupting actions of Alzheimer's disease A β in vivo. *Philos Trans R Soc Lond B Biol Sci.* 2014;369(1633):20130147.
- Yun SH, Gamkrelidze G, Stine WB, Sullivan PM, Pasternak JF, Ladu MJ, et al. Amyloid-beta1-42 reduces neuronal excitability in mouse dentate gyrus. *Neurosci Lett.* 2006;403(1-2):162-5.
- Pocernich CB, Butterfield DA. Acrolein inhibits NADH-linked mitochondrial enzyme activity: implications for Alzheimer's disease. *Neurotox Res.* 2003;5(7):515-20.
- Matousek SB, Ghosh S, Shaftel SS, Kyrkanides S, Olschowka JA, O'Banion MK. Chronic IL-1 β -mediated neuroinflammation mitigates amyloid pathology in a mouse model of Alzheimer's disease without inducing overt neurodegeneration. *J Neuroimmune Pharmacol.* 2012;7(1):156-64.
- Zussy C, Brureau A, Delair B, Marchal S, Keller E, Ixart G, et al. Time-course and regional analyses of the physiopathological changes induced after cerebral injection of an amyloid beta fragment in rats. *Am J Pathol.* 2011;179(1):315-34.
- Yamaguchi Y, Kawashima S. Effects of amyloid-beta-(25-35) on passive avoidance, radial-arm maze learning and choline acetyltransferase activity in the rat. *Eur J Pharmacol.* 2001;412(3):265-72.
- Mogensen J, Pedersen TK, Holm S, Bang LE. Prefrontal cortical mediation of rats' place learning in a modified water maze. *Brain Res Bull.* 1995;38(5):425-34.
- Sepulveda FJ, Fierro H, Fernandez E, Castillo C, Peoples RW, Opazo C, et al. Nature of the neurotoxic membrane actions of amyloid-beta on hippocampal neurons in Alzheimer's disease. *Neurobiol Aging.* 2014;35(3):472-81.
- Stepanichev M, Lazareva N, Tukhbatova G, Salozhin S, Gulyaeva N. Transient disturbances in contextual fear memory induced by A β (25-35) in rats are accompanied by cholinergic dysfunction. *Behav Brain Res.* 2014;259:152-7.
- Asi SS, Farhadi HM, Naghdi N, Choopani S, Samzadeh-Kermani A, Mehdizadeh M. Non-acute effects of different doses of 3, 4-methylenedioxymethamphetamine on spatial memory in the morris water maze in Sprague-Dawley male rats. *Neural Regen Res.* 2011;6:1715.
- Nitta A, Fukuta T, Hasegawa T, Nabeshima T. Continuous infusion of beta-amyloid protein into the rat cerebral ventricle induces learning impairment and neuronal and morphological degeneration. *Jpn J Pharmacol.* 1997;73(1):51-7.
- Maurice T, Lockhart BP, Privat A. Amnesia induced in mice by centrally administered beta-amyloid peptides involves cholinergic dysfunction. *Brain Res.* 1996;706(2):181-93.
- Harkany T, O'Mahony S, Kelly JP, Soos K, Toro I, Penke B, et al. Beta-amyloid(Phe(SO₃H)₂₄)₂₅₋₃₅ in rat nucleus basalis induces behavioral dysfunctions, impairs learning and memory and disrupts cortical cholinergic innervation. *Behav Brain Res.* 1998;90(2):133-45.
- Selkoe DJ. Alzheimer's disease is a synaptic failure. *Science.* 2002;298(5594):789-91.
- Trubetskaya VV, Stepanichev MY, Onufriev MV, Lazareva NA, Markevich VA, Gulyaeva NV. Administration of aggregated beta-amyloid peptide (25-35) induces changes in long-term potentiation in the hippocampus in vivo. *Neurosci Behav Physiol.* 2003;33(2):95-8.
- Abramov AY, Canevari L, Duchen MR. Beta-amyloid peptides in-

- duce mitochondrial dysfunction and oxidative stress in astrocytes and death of neurons through activation of NADPH oxidase. *J Neurosci*. 2004;**24**(2):565-75.
22. Quintanilla RA, Orellana DI, Gonzalez-Billault C, Maccioni RB. Interleukin-6 induces Alzheimer-type phosphorylation of tau protein by deregulating the cdk5/p35 pathway. *Exp Cell Res*. 2004;**295**(1):245-57.
 23. Takeuchi H, Jin S, Wang J, Zhang G, Kawanokuchi J, Kuno R, et al. Tumor necrosis factor-alpha induces neurotoxicity via glutamate release from hemichannels of activated microglia in an autocrine manner. *J Biol Chem*. 2006;**281**(30):21362-8.
 24. Viegas CM, Tonin AM, Zanatta A, Seminotti B, Busanello EN, Fernandes CG, et al. Impairment of brain redox homeostasis caused by the major metabolites accumulating in hyperornithinemia-hyperammonemia-homocitrullinuria syndrome in vivo. *Metab Brain Dis*. 2012;**27**(4):521-30.
 25. Azad N, Rasoolijazi H, Joghataie MT, Soleimani S. Neuroprotective effects of carnolic Acid in an experimental model of Alzheimer's disease in rats. *Cell J*. 2011;**13**(1):39-44.
 26. Marzban M, Asl SS, Huseini HF, Tondar M, Choopani S, Mehdizadeh M. Effects of butternut squash extract on dentate gyrus cell proliferation and spatial learning in male adult rats. *Neural Reg Res*. 2011;**6**(24).
 27. Mehdizadeh M, Dabaghian F, Nejhad A, Fallah-Huseini H, Choopani S, Shekarriz N, et al. Zingiber Officinale Alters 3,4-methylenedioxymethamphetamine-Induced Neurotoxicity in Rat Brain. *Cell J*. 2012;**14**(3):177-84.
 28. Nezhadi A, Ghazi F, Rassoli H, Bakhtiari M, Ataiy Z, Soleimani S, et al. BMSC and CoQ10 improve behavioural recovery and histological outcome in rat model of Parkinson's disease. *Pathophysiology*. 2011;**18**(4):317-24.
 29. Rasoolijazi H, Azad N, Joghataie MT, Kerdari M, Nikbakht F, Soleimani M. The protective role of carnolic acid against beta-amyloid toxicity in rats. *ScientificWorldJournal*. 2013;**2013**:917082.
 30. Soleimani AS, Pourheydar B, Dabaghian F, Nezhadi A, Rooiantan A, Mehdizadeh M. Ecstasy-Induced Caspase Expression Alters Following Ginger Treatment. *Basic Clinic Neuros*. 2013;**4**(4):51-5.
 31. Peng QL, Buz'Zard AR, Lau BH. Pycnogenol protects neurons from amyloid-beta peptide-induced apoptosis. *Brain Res Mol Brain Res*. 2002;**104**(1):55-65.